



# SEPARATION REPORT

## GFC separation of water-soluble polymers with TSKgel PW-type columns (2) — using the PW<sub>XL</sub> series —

### — Contents —

	Page
1. Introduction	1
2. Calibration curves, theoretical plate numbers and separation range for each grade in the PW <sub>XL</sub> series	1
3. Comparison of performance of PW <sub>XL</sub> series and PW series	3
4. Effect of elution conditions on resolution of the PW <sub>XL</sub> series	4
Effect of flow rate	4
Effect of temperature	4
Effect of sample concentration	4
Effect of sample injection volume	5
5. Relationship between theoretical plate number and resolution factor	6
6. Dependence of $M_w/M_n$ and $M_z/M_w$ on theoretical plate number and sample concentration in GPC	6
7. Analyzing molecular mass by GPC/LALLS (low-angle laser light scattering detector) using TSKgel GMPW <sub>XL</sub>	8
8. Dependence of $M_n$ , $M_w$ , and $M_z$ on theoretical plate number with GPC/LALLS	9
9. Selection of Eluents for GPC separation of water-soluble polymers	10
10. Conclusion	13

## 1. Introduction

During the last dozen years, high-performance GPC and GFC have gained widespread use due to their ability to facilitate separation, good reproducibility, and short separation time.

As shown in Table 1, which indicates the years in which various packing materials for GPC and GFC were developed, Tosoh has led the world in developing high-performance GPC and GFC columns for both organic solvents and aqueous solvent systems.

Now, at a time when demand is rising for faster, more sensitive, and labor-saving analytical equipment, Tosoh is developing high-performance high resolution columns for both organic solvent systems and aqueous solvent systems.

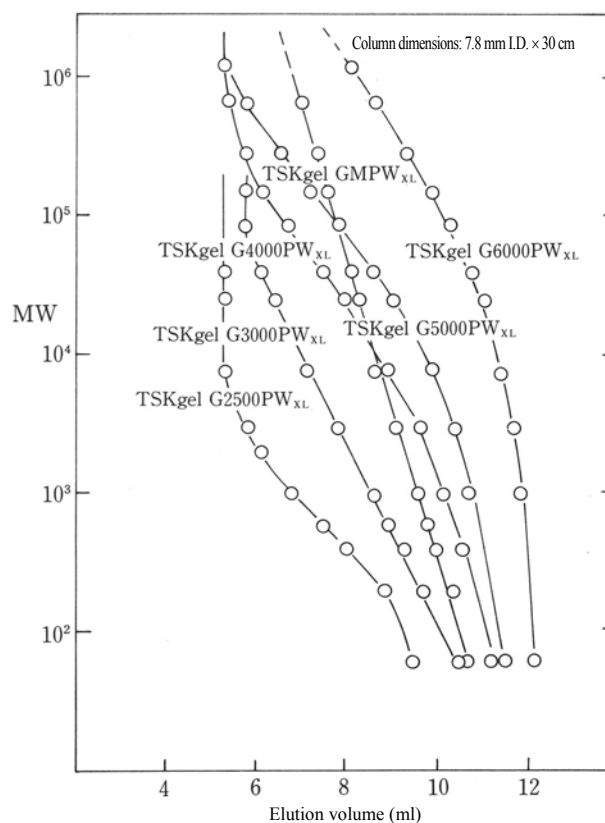
This report discusses the separation performance of the high-performance PW<sub>XL</sub> series of GPC columns for use in aqueous systems, and presents applications for using the PW<sub>XL</sub> series.

**Table 1 History of development of GPC and GFC packing materials**

		Historical background
1970	Development of S- and H-type GPC packing materials for organic solvent systems	Petrochemical industry is flourishing
1975	Development of SW- and PW-type GFC packing materials for aqueous solvent systems	Oil shock Growth of high value-added industries
1980	Development of TOYOPEARL packing materials for preparative GFC Development of packing materials for GPC and GFC with high theoretical plate numbers	Analytical equipment becoming faster, more sensitive, and more laborsaving

## 2. Calibration curves, theoretical plate numbers and separation range for each grade in the PW<sub>XL</sub> series

Figure 1 shows calibration curves for each column in the PW<sub>XL</sub> series using a standard polyethylene oxide (PEO) SE series sold by Tosoh. TSKgel GMPW<sub>XL</sub> has been added which is absent from the conventional PW series of columns. The TSKgel GMPW<sub>XL</sub> is a series of columns designed so that when several grades of columns with different pore sizes are combined, the calibration curve will become linear over a wide range of molecular masses, making this system suitable for analyzing molecular mass and molecular mass distributions. The TSKgel G2500PW<sub>XL</sub> has also been added. In this grade, ion specificity is vastly improved over that of the conventional TSKgel G2000PW column, while the resolution is maintained (for details see Separation Report No. 37).



Sample: Standard polyethylene oxide

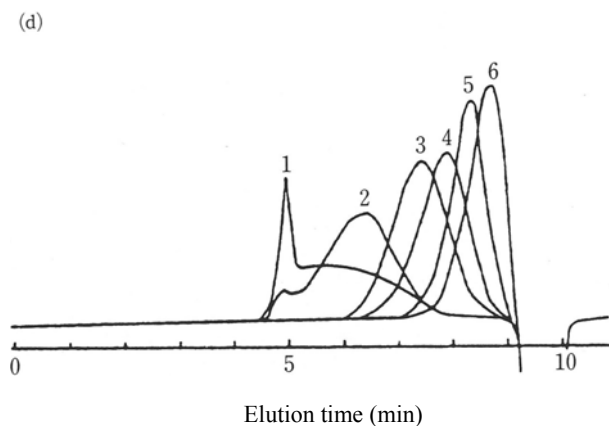
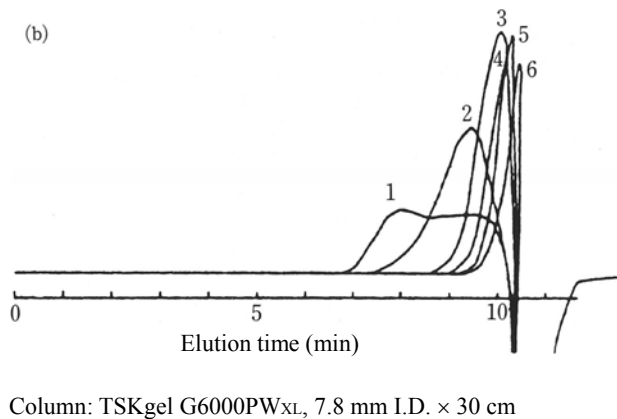
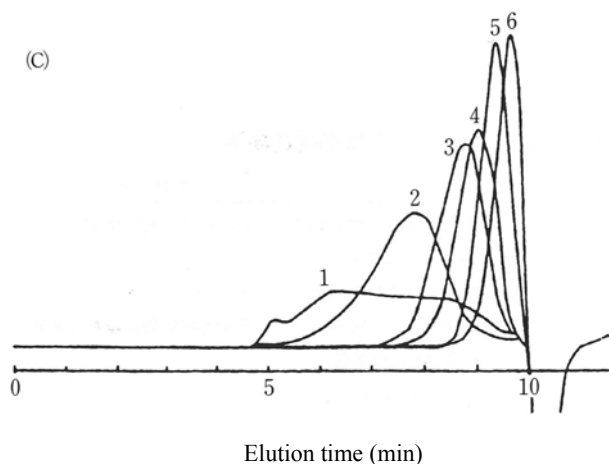
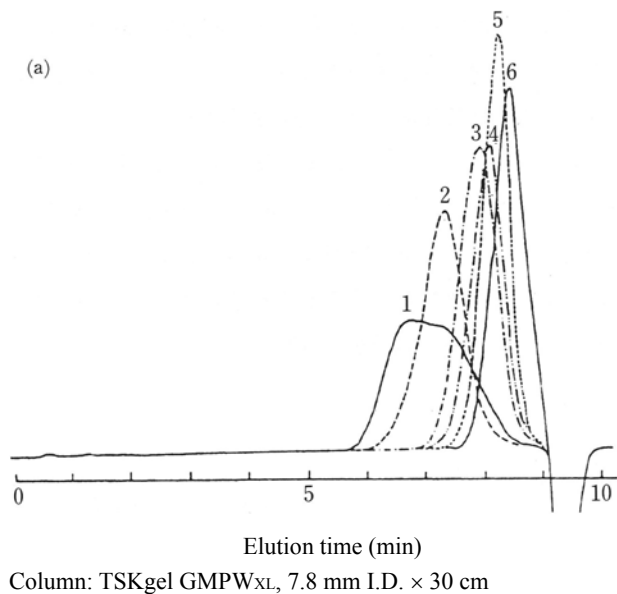
**Figure 1 Calibration curves for the PW<sub>XL</sub> series**

Table 2 shows the name, particle size, theoretical plate number and molecular mass exclusion limit of each grade. Figures 2a through 2d are chromatograms produced by separating dextran using the TSKgel GMPW<sub>XL</sub>, TSKgel G6000PW<sub>XL</sub>, TSKgel G5000PW<sub>XL</sub>, and TSKgel G4000PW<sub>XL</sub>. Consult Figures 1 and 2 and Table 2 to choose the optimum column system for the sample to be analyzed.

**Table 2 TSKgel PW<sub>XL</sub> Series**

Name	Particle size	Theoretical plate number	Exclusion limit
G2500PW <sub>XL</sub>	6µm	14000TP	10,000Da
G3000PW <sub>XL</sub>	6	14000	100,000
G4000PW <sub>XL</sub>	10	10000	1,000,000
G5000PW <sub>XL</sub>	10	10000	4,000,000
G6000PW <sub>XL</sub>	13	7000	—
GMPW <sub>XL</sub>	13	7000	—
G-Oligo-PW	6	14000	10,000
G-DNA-PW	10	—	—

(Size of each column: 7.8 mm I.D. × 30 cm)



**Figure 2 Separation of dextran**

Injection volume: 100 µL (1.0 g/L)

Temperature: 40 °C

Eluent: 0.2 mol/L phosphate buffer (pH 6.9)

Flow rate: 1.0 mL/min

Detection: RI

Samples

1	Dextran	T-2000
2		T-500
3		T-70
4		T-40
5		T-20
6		T-10

### 3. Comparison of performance of PW<sub>XL</sub> series and PW series

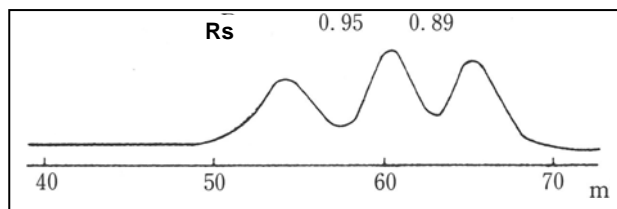
Figures 3 and 4 are chromatograms comparing the performance of the PW and PW<sub>XL</sub> series of columns using a TSKgel G6000PW<sub>(XL)</sub> + TSKgel G3000PW<sub>(XL)</sub> system of columns.

Figure 3 compares elution profiles and resolution ( $R_s$ ) produced using a mixture of polyethylene oxide standards (SE

150:  $M_w = 1.20 \times 10^6$ ; SE 30:  $M_w = 2.8 \times 10^6$ ; SE 8:  $M_w = 7.3 \times 10^4$ ) as the sample. Figure 4 compares elution profiles produced using a mixture of pullulan standards (P-400:  $M_w = 33.8 \times 10^4$ ; P-50:  $M_w = 4.67 \times 10^4$ ) as the sample. As columns in the PW<sub>XL</sub> series are half as long as the 60-cm columns in the PW series, separation is completed in half the time, while resolution shows equivalent or higher resolution than results produced with the PW series.

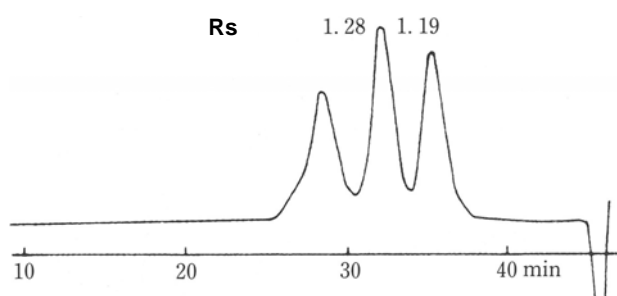
PW Series

Column size: 7.5 mm I.D.  $\times$  60 cm  $\times$  2



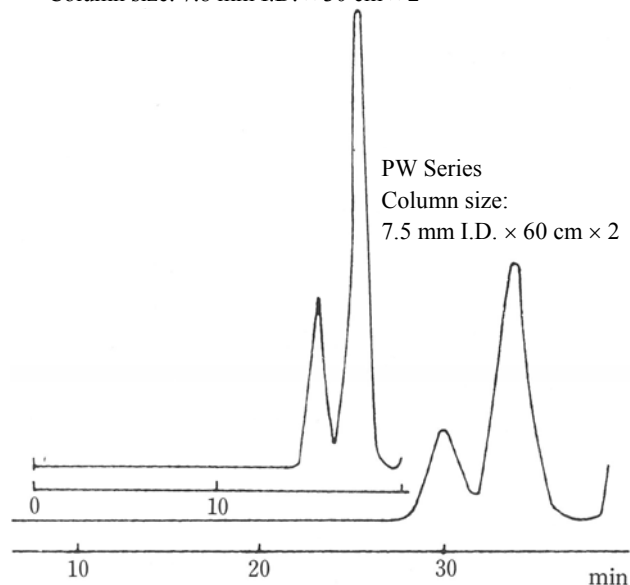
PW<sub>XL</sub> Series

Column size: 7.8 mm I.D.  $\times$  30 cm  $\times$  2



PW<sub>XL</sub> Series

Column size: 7.8 mm I.D.  $\times$  30 cm  $\times$  2



**Figure 3 Comparison of PW and PW<sub>XL</sub> Series (1)**

Sample: Polyethylene oxide, SE150-SE30-SE8  
 Injection volume: 100  $\mu$ L (0.4 g/L)  
 Column: TSKgel G6000 PW + TSKgel G3000 PW  
 Temperature: 50  $^{\circ}$ C  
 Eluent: 0.1 mol/L NaCl  
 Flow rate: 0.5 mL/min

**Figure 4 Comparison of PW and PW<sub>XL</sub> Series (2)**

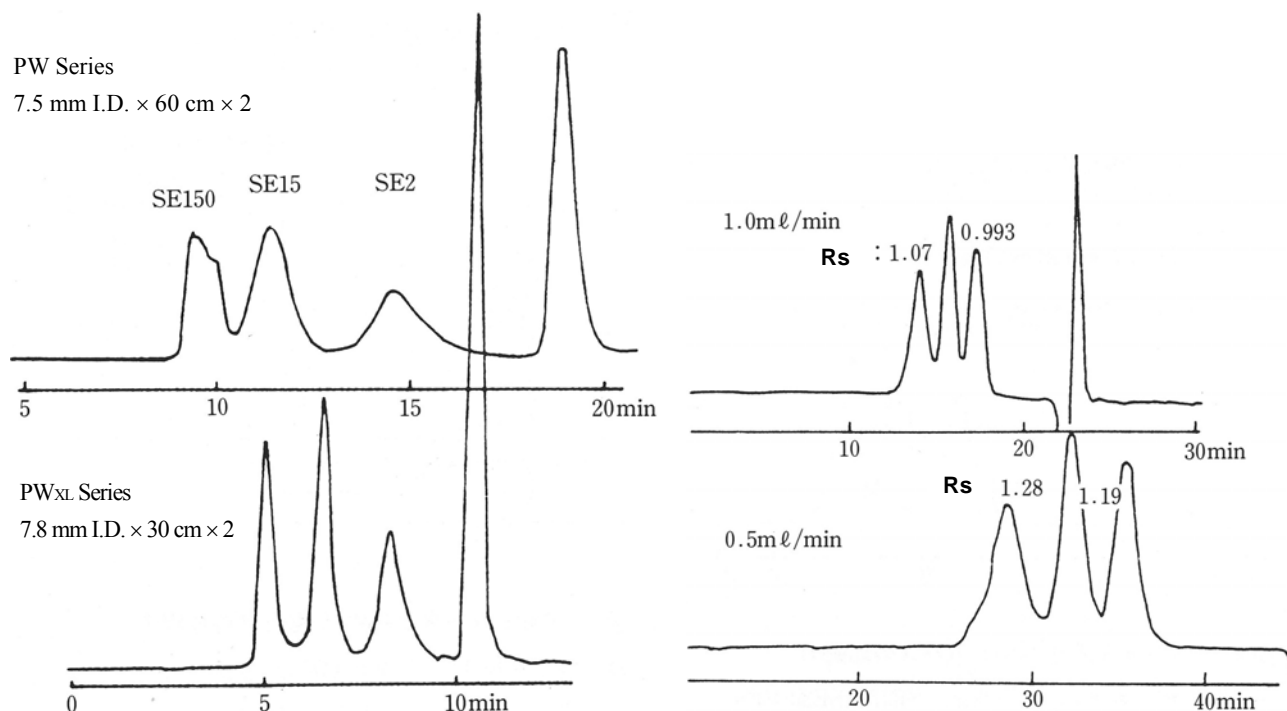
Sample: Pullulan, P400-P50  
 Injection volume: 100  $\mu$ L (0.4, 1.2 g/L)  
 Column: TSKgel G6000 PW + TSKgel G3000 PW  
 Temperature: 50  $^{\circ}$ C  
 Eluent: 0.1 mol/L NaCl  
 Flow rate: 1.0 mL/min

Figure 5 shows the results of a comparison of the performance of PW<sub>XL</sub> series with the PW series in a system using a TSKgel G4000 PW column by itself. A mixture of polyethylene oxide standards (SE 150:  $M_w = 1.20 \times 10^6$ ; SE 15:  $M_w = 15 \times 10^4$ ; SE 2:  $M_w = 2.5 \times 10^4$ ) was used as the sample. Results were similar to those shown in Figures 3 and 4 were obtained.

#### 4. Effect of elution conditions on resolution of the PW<sub>XL</sub> series

##### 1) Effect of flow rate

Figure 6 compares elution profiles at flow rates of 1.0 mL/min and 0.5 mL/min. Resolution ( $R_s$ ) between the peaks are noted on the chromatographs. Results show that resolution improves as the flow rate decreases.



**Figure 5 Comparison of PW and PW<sub>XL</sub> Series (3)**

Sample: Standard polyethylene oxide  
 Injection volume: 100  $\mu$ L  
 Column: TSKgel G4000 PW  
 Temperature: 50  $^{\circ}$ C  
 Eluent: 0.1 mol/L NaCl  
 Flow rate: 1.0 mL/min

##### 2) Effect of temperature

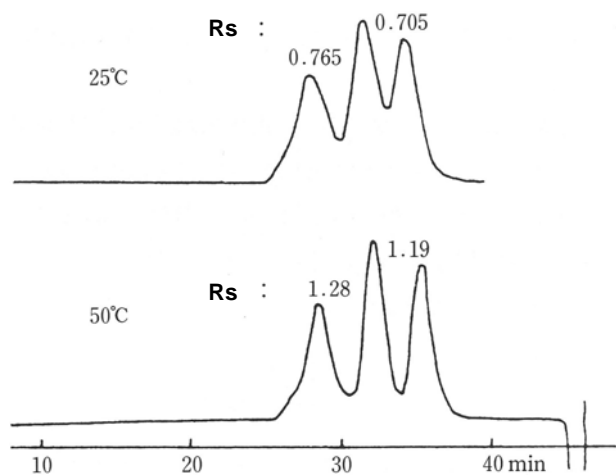
Figure 7 shows the results of a comparison of the elution profiles occurring when columns were placed in a column oven at 25  $^{\circ}$ C and 50  $^{\circ}$ C, at a constant flow rate (0.5 mL/min). Increasing the temperature causes the resolution between the peaks to increase significantly.

##### 3) Effect of sample concentration

Figure 8 compares elution profiles produced when the sample concentration of each component was 0.4 g/L versus 1.6 g/L. Elution profiles are very sensitive to the sample concentration and change dramatically as shown in Figure 8. In general, separation improves as the concentration of the sample decreases.

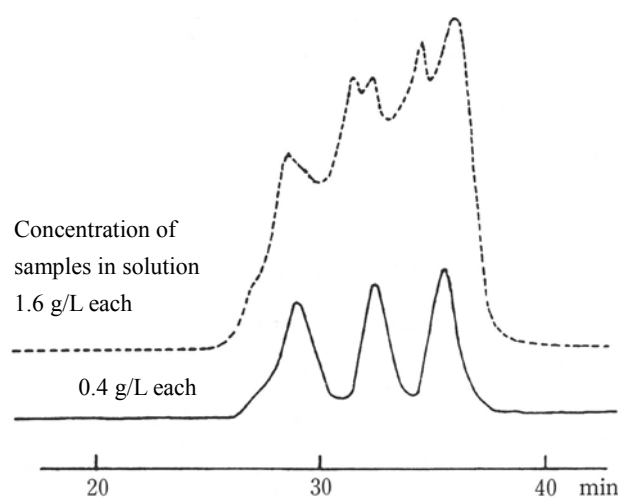
**Figure 6 Effect of flow rate**

Sample: Standard polyethylene oxide, SE150-SE30-SE8  
 Injection volume: 100  $\mu$ L (0.4 g/L)  
 Columns: TSKgel G6000PW<sub>XL</sub> + TSKgel 3000 PW<sub>XL</sub>  
 7.8 mm I.D. x 30 cm x 2  
 Temperature: 50  $^{\circ}$ C  
 Eluent: 0.1 mol/L NaCl



**Figure 7 Effect of temperature**

Sample: Standard polyethylene oxide,  
SE150-SE30-SE8  
Injection volume: 100  $\mu\text{L}$  (0.4 g/L)  
Columns: TSKgel G6000PW<sub>XL</sub> + TSKgel G3000PW<sub>XL</sub>  
7.8 mm I.D.  $\times$  30 cm  $\times$  2  
Eluent: 0.1 mol/L NaCl  
Flow rate: 0.5 mL/min

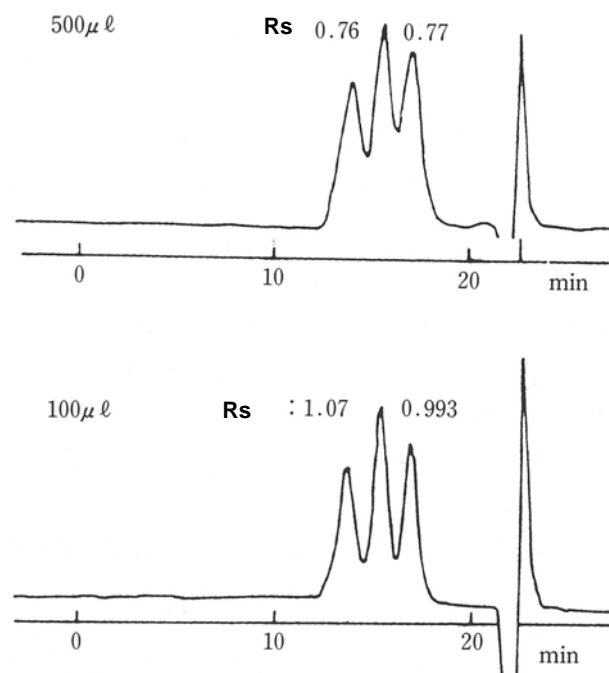


**Figure 8 Effect of sample concentration in solution**

Sample: Standard polyethylene oxide,  
SE150-SE30-SE8  
Injection volume: 100  $\mu\text{L}$  (0.4 g/L)  
Columns: TSKgel G6000PW<sub>XL</sub> + TSKgel G3000PW<sub>XL</sub>  
7.8 mm I.D.  $\times$  30 cm  $\times$  2  
Temperature: 50°C  
Eluent: 0.1 mol/L NaCl  
Flow rate: 0.5 mL/min

#### 4) Effect of sample injection volume

Figure 9 compares the elution profiles produced with sample injection volumes of 100  $\mu\text{L}$  and 500  $\mu\text{L}$ . At the same sample load, the degree of change in  $R_s$  is less when the injection volume is increased without changing the concentration of the sample in the solution, compared to when the concentration of the sample in the solution is increased without changing the injection volume (see Figures 8 and 9). When analysis sensitivity is low, increasing the injection volume improves precision. For reference, Figure 10 illustrates the effect of injection volume on elution profiles with the PW series. The PW<sub>XL</sub> series is slightly more sensitive to injection volume than the PW series.



**Figure 9 Effect of sample injection volume (1)**

Sample: Standard polyethylene oxide,  
SE150-SE30-SE8  
Sample concentration: 0.4 g/L each  
Columns: TSKgel G6000PW<sub>XL</sub> + TSKgel G3000PW<sub>XL</sub>  
7.8 mm I.D.  $\times$  30 cm  $\times$  2  
Temperature: 50°C  
Eluent: 0.1 mol/L NaCl  
Flow rate: 1.0 mL/min

## 5. Relationship between theoretical plate number and resolution (Rs)

In high-performance liquid chromatography, the separation between samples is expressed by Equation (1).

$$R_s = \frac{1}{4} \sqrt{N} \left( \frac{\alpha - 1}{\alpha} \right) \cdot \left( \frac{K'}{K' + 1} \right) \dots\dots\dots(1)$$

Here,  $R_s$  represents the resolution of a sample, with better separation indicated by larger values.  $N$  expresses the theoretical plate number;  $\alpha$  as the separation factor of the sample and  $K'$  as the capacity ratio are calculated using Equations (2) and (3), respectively.

$$\alpha = \frac{V_{R2} - V_0}{V_{R1} - V_0} \dots\dots\dots(2)$$

$$K' = \frac{V_R - V_0}{V_0} \dots\dots\dots(3)$$

(See Figure 11 for  $V_R$  and  $V_0$ )

The method for calculating the resolution  $R_s$  is shown in Figure 11.

In GPC, because the interaction between the sample and the packing material is not used,  $\alpha$  and  $K'$  are constants that are unrelated to the length of the column.

Thus Equation (1) becomes:

$$R_s = K' \sqrt{N} \dots\dots\dots(1') \quad K: \text{constant value}$$

A simple way to increase the number of theoretical plates is to lengthen the column.

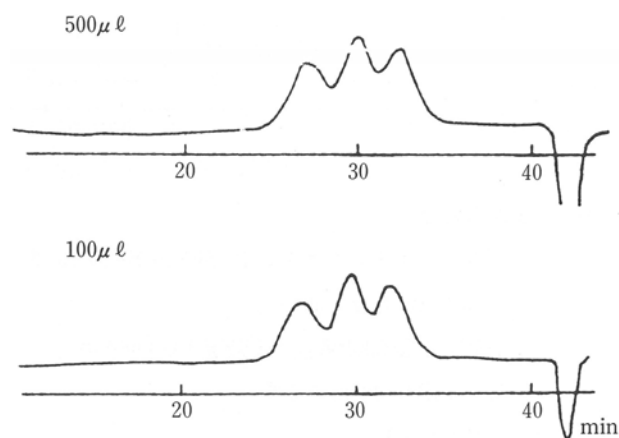
Figure 12 shows changes in the elution profiles for a mixed solution of standard polyethylene oxides produced when the length of the column is changed. Separation increases as the length of the column increases. Table 3 shows theoretical plate numbers  $N$  for SE15 at various column lengths as well as calculations of the  $R_s$  between peaks SE15 and SE2.  $R$  and  $\sqrt{N}$  values are plotted in Figure 13. A very good linear relationship is achieved, satisfying the relationship expressed by equation (1').

## 6. Dependence of $M_w/M_n$ and $M_z/M_w$ on theoretical plate number and sample concentration in GPC

Using standard polyethylene oxide SE15, pullulan P50 and P400, and dextran T70 as samples, changes in  $M_w/M_n$  and  $M_z/M_w$  values at varying column lengths were investigated using TSKgel G4000PW<sub>XL</sub> in the column. The results are shown in Figure 14. Values for dextran, pullulan and polyethylene oxide tended to near constant a level at a column length of  $\geq 120$  cm.

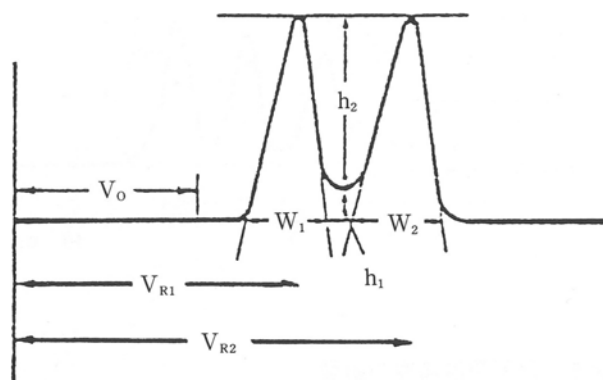
Figure 15 shows changes in  $M_w/M_n$  and  $M_z/M_w$  values produced when the concentration of the sample in solution was varied at a fixed column length of 30 cm.  $M_w/M_n$  and  $M_z/M_w$  values became constant in all solutions at a concentration range of 0.05-0.2% (injection volume: 100  $\mu$ L).

For each of these samples,  $M_w/M_n$  and  $M_z/M_w$  values were not as sensitive to resolution as the theoretical plate number or resolution. However, to obtain accurate  $M_w/M_n$  and  $M_z/M_w$  values, a column length of around 120 cm is necessary when using TSKgel G4000PW<sub>XL</sub>.



**Figure 10 Effect of sample injection volume (2)**

Sample: Standard polyethylene oxide, SE150-SE30-SE8  
 Sample solution concentration: 0.4 g/L each  
 Columns: TSKgel G6000PW + TSKgel G3000PW  
 7.5 mm I.D.  $\times$  60 cm  $\times$  2  
 Temperature: 50°C  
 Eluent: 0.1 mol/L NaCl  
 Flow rate: 1.0 mL/min

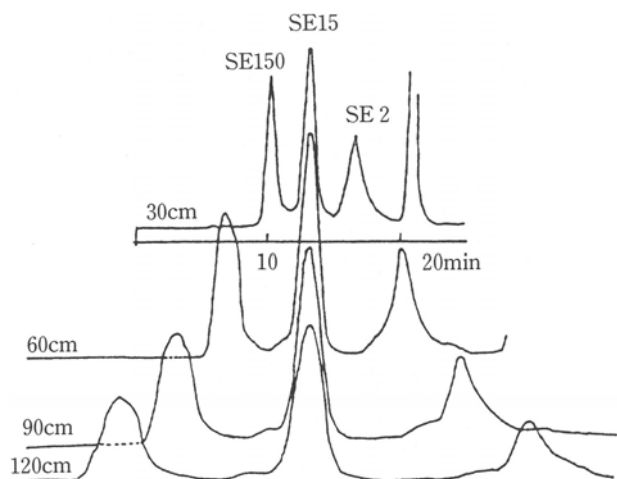


**Figure 11 Method for calculating resolution factor**

$$R = \frac{2(V_{R2} - V_{R1})}{W_1 + W_2} \dots\dots\dots(a) \text{ or}$$

$$R = \frac{h_2}{h_1 + h_2} \times 100 \dots\dots\dots(b)$$

In (a), the value increases as separation improves, while the value in (b) will become 100% when separation is complete.



**Figure 12** Dependence of changes in separation on column length

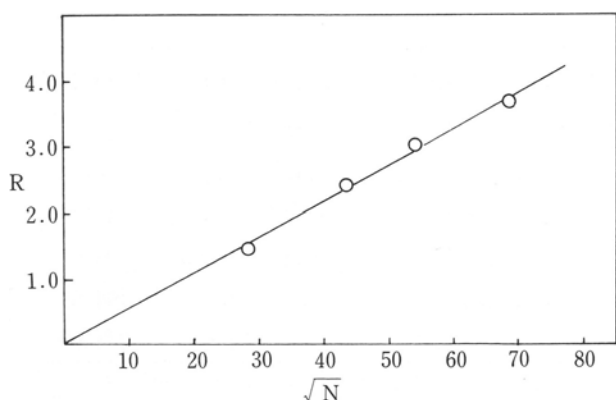
Sample: Standard polyethylene oxide,  
SE150-SE15-SE2  
Injection volume: 100  $\mu$ L (0.4 g/L each)  
Column: TSKgel G4000PW<sub>XL</sub>  
7.8 mm I.D.  
Temperature: 50°C  
Eluent: 0.1 mol/L NaCl  
Flow rate: 0.5 mL/min

**Table 3** Theoretical plate number and resolution factor (1)

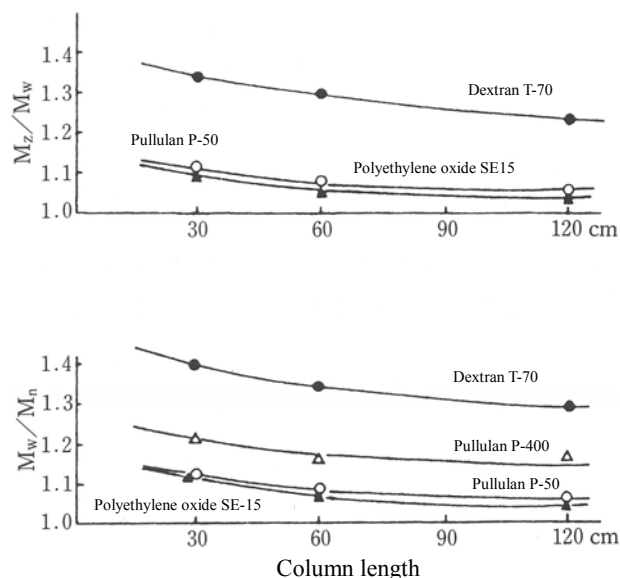
Column length (cm)	N*	$\sqrt{N}$	R**
30	800	28.3	1.476
60	1900	43.6	2.444
90	2930	54.1	3.04
120	4675	68.4	3.69

\*N: Theoretical plate number calculated from peak for SE15.

\*\*R: Resolution factor between peaks for SE15 and SE2.

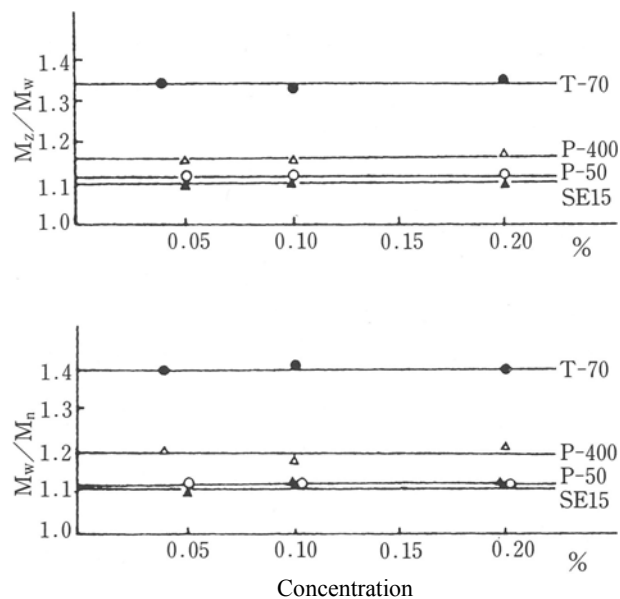


**Figure 13** Theoretical plate number and resolution (2)



**Figure 14** Dependence on theoretical plate number

Column: TSKgel G4000PW<sub>XL</sub>  
7.8 mm I.D.  
Temperature: 50°C  
Eluent: 0.1 mol/L NaCl  
Flow rate: 1.0 mL/min  
Injection volume: 100  $\mu$ L (1.0 g/L each)



**Figure 15** Dependence on concentration of sample solution

Column: TSKgel G4000PW<sub>XL</sub>  
7.8 mm I.D.  $\times$  30 cm  
Temperature: 50 °C  
Eluent: 0.1 mol/L NaCl  
Flow rate: 1.0 mL/min  
Injection volume: 100  $\mu$ L



However, currently the most problematic area of GPC with aqueous systems is that it is not possible to obtain a series of standard samples with a molecular mass of 1 million or higher. This often becomes inconvenient for calculating  $M_n$ ,  $M_w$  and  $M_z$ , and development of a series of standard samples with high molecular mass is needed for use in aqueous systems.

## 7. Analyzing molecular mass by GPC/LALLS (low-angle laser light scattering detector) using TSKgel GMPW<sub>XL</sub>

1) Analyzing weight-average molecular mass  $M_w$  of pullulan standard sample

Pullulan standard samples with a narrow molecular mass distribution are commercially available. Accordingly, the molecular mass of pullulan was subjected to GPC/LALLS separation using a TSKgel GMPW<sub>XL</sub> column. Figure 16 shows the GPC/LALLS chromatogram produced.

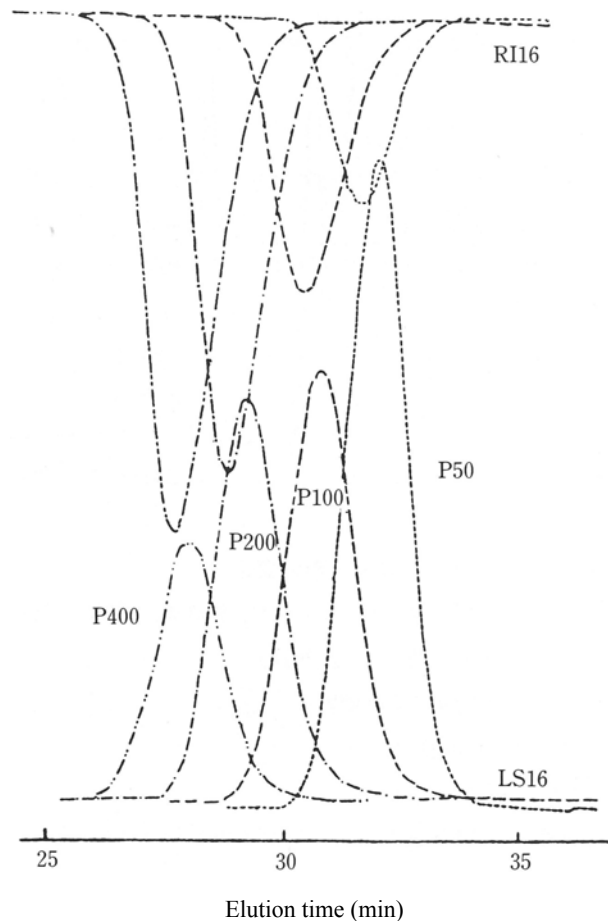
Table 4 shows the results of separation for each grade of pullulan. Human serum albumin ( $M=66,000$ ) was used as the primary standard, with pullulan P100 as the secondary standard. For pullulan, molecular masses were determined by Professor Kawahara et al., by sedimentation equilibrium. Molecular masses were also determined by Professor Takagi et al., by GPC/LALLS, and a comparison found these results were very consistent.

2) Measuring the weight-average molecular mass  $M_w$  of standard sodium polystyrene sulfonate (PNaSS)

Standard sodium polystyrene sulfonate is commercially available from the American company, Pressure Chemical, as a standard polymer electrolyte with a narrow molecular mass distribution.

GPC separation of standard sodium polystyrene sulfonate requires the addition of at least 10 % acetonitrile or methanol to the eluent. Figure 18 shows chromatograms produced when standard sodium polystyrene sulfonate was analyzed using TSKgel GMP<sub>XL</sub>. Peak shapes on a chromatogram of standard sodium polystyrene sulfonate produced with 10 % added acetonitrile were essentially the same as when 20 % acetonitrile was added.

Table 5 shows the results of a study of the differences in the apparent molecular mass  $M_w$  of each sample with varying Eluent compositions. Small changes in the concentration of inorganic salt in the solvent or in the composition of an organic solvent caused only very slight changes in apparent  $M_w$ . However, the values resulting from analyses in each case were slightly higher than the nominal value provided by Pressure Chemical. Various explanations for this have been considered, including the complexity of the solvent composition, the inability of this method to handle light scattering in two-component solution, and the hygroscopic properties of the samples. This problem will be a subject of future research in GPC/LALLS.



**Figure 16 Chromatogram of pullulan by GPC/LALLS**

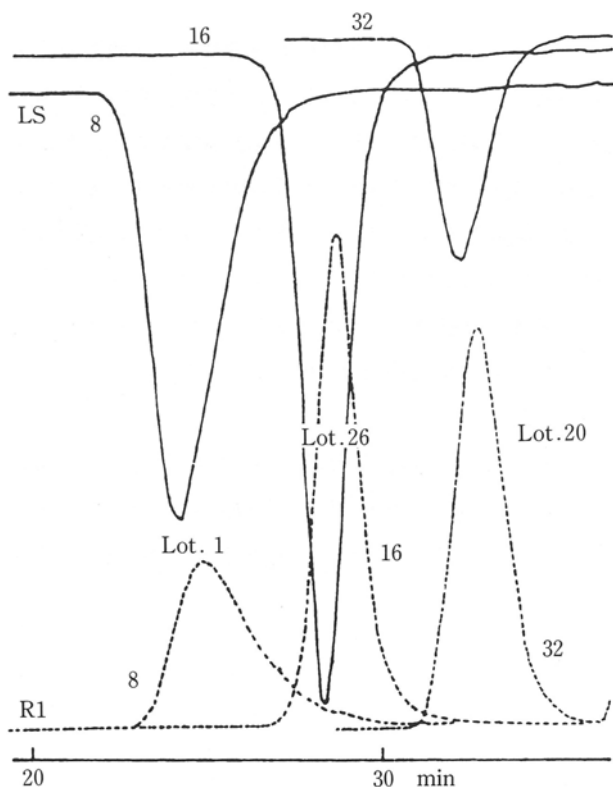
Sample: Pullulan  
 Injection volume: 500  $\mu$ L  
 Column: TSKgel GMPW<sub>XL</sub>  
 7.8 mm I.D.  $\times$  120 cm  
 Temperature: 40  $^{\circ}$ C  
 Eluent: 0.1 mol/L NaCl  
 Flow rate: 1.0 mL/min

**Table 4 Results of pullulan molecular mass analysis**

Grade of pullulan	$M_w$	Rf.1	Rf.2
P-10	10,100	10,400	12,000
P-20	18,600	18,200	20,800
P-50	45,700	45,500	46,700
P-100	98,800	100,000	95,400
P-200	177,500	187,000	194,000
P-400	334,000	348,000	338,000

Rf.1 Data from Professor Takagi et al.

Rf.2 Data from sedimentation equilibrium by Professor Kawahara et al.



**Figure 18 Separation of standard sodium polystyrene sulfonate**

Sample: Standard sodium polystyrene sulfonate  
 Injection volume: 500  $\mu$ L  
 Column: TSKgel GMPW<sub>XL</sub>  
 7.8 mm I.D.  $\times$  30 cm  $\times$  4  
 Temperature: 40  $^{\circ}$ C  
 Eluent: 10 % acetonitrile/0.2 mol/L Na<sub>2</sub>SO<sub>4</sub>  
 Flow rate: 1.0 mL/min

## 8. Dependence of $M_n$ , $M_w$ , and $M_z$ on theoretical plate number with GPC/LALLS

Table 6 shows changes in  $M_n$ ,  $M_w$ , and  $M_z$  when analysis was performed by GPC/LALLS at varying TSKgel GMPW<sub>XL</sub> column lengths, using dextran T500 as the sample. Values in parentheses indicate values without performing function approximation. (The necessity and validity of functional approximation is published in a separate report). Although slight,  $M_n$ ,  $M_w$ , and  $M_z$  values do depend on the theoretical plate number. (As in normal GPC, a column with a certain capacity for high-resolution separation is necessary to evaluate  $M_n$  and  $M_z$ ).

**Table 6 Analysis of weight-average molecular mass by GPC/LALLS**

Column length (cm)	$M_n \times 10^{-4}$	$M_w \times 10^{-4}$	$M_z \times 10^{-4}$
60	20.6 (19.7)	43.2 (44.2)	123.4 (139.0)
120	19.5 (20.2)	44.1 (44.1)	139.8 (140.5)
240	19.3 (19.1)	45.4 (45.1)	143.5 (139.0)

Values in parentheses were calculated without functional approximation.

Sample: Dextran T500  
 Column: TSKgel GMPW<sub>XL</sub>  
 Solution: 0.1 mol/L NaCl

**Table 5 Analysis of weight-average molecular mass of standard sodium polystyrene sulfonate by GPC/LALLS**

Lot. No.	Solvent a	Apparent $M_w$			Nominal value
		b	c		
#1	1210000	1210000	1290000		1060000
#16	794000	787000	821000		690000
#26	202500	202000	210000		177000
#20	19400	21000			16000

### Solvents

- a: 10 % CH<sub>3</sub>CN/0.2 mol/L Na<sub>2</sub>SO<sub>4</sub>
- b: 20 % CH<sub>3</sub>CN/0.2 mol/L Na<sub>2</sub>SO<sub>4</sub>
- c: 10 % CH<sub>3</sub>CN/0.4 mol/L Na<sub>2</sub>SO<sub>4</sub>

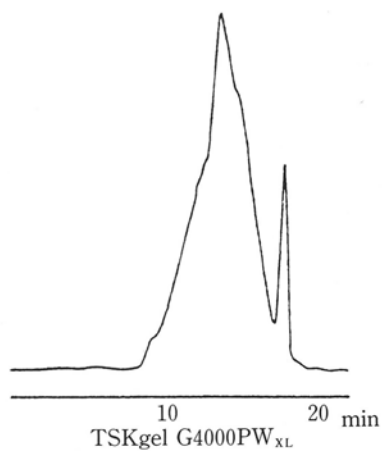
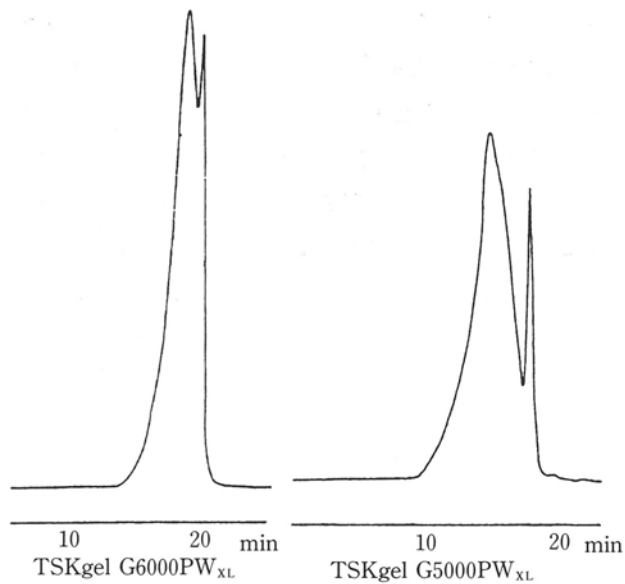
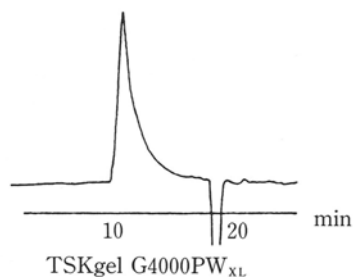
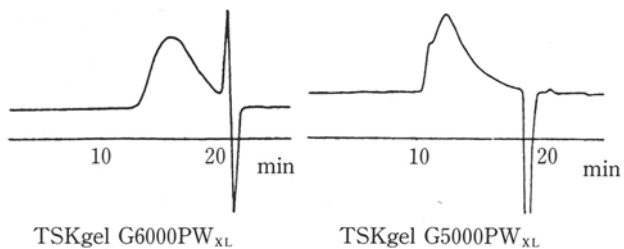
## 9. Selection of solvents for GPC separation of water-soluble polymers

Table 7 shows a table listing various types of representative water-soluble polymers. Although various types of water-soluble polymers exist, most samples can be analyzed by preparing several solvents for GPC use as shown in Table 8. However, analyzing viscose is not currently easy, as this substance can only be dissolved in a highly alkaline aqueous solution, or polymers that contain several aromatic rings (in monomer units).

To date, the separation of polycations has been considered to be difficult due to the fact that trace quantities of carboxyl groups are present in the gel. However, this problem can be solved by making the solvent acidic, inhibiting carboxyl group disassociation, and increasing the ionic strength. One problem is that, in our experience, if polyanions are analyzed after polycation separation has been performed, polyanions, which should be easy to analyze, show signs of adsorption. As this can cause various problems in separation, setting aside columns that are used only for polycation separation is preferable if possible. Several GPC applications of water-soluble polymers are shown in Figures 19-24.

**Table 7** Types of water-soluble polymers

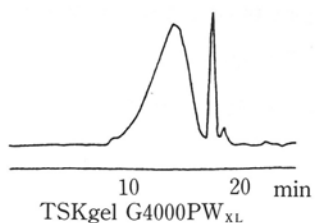
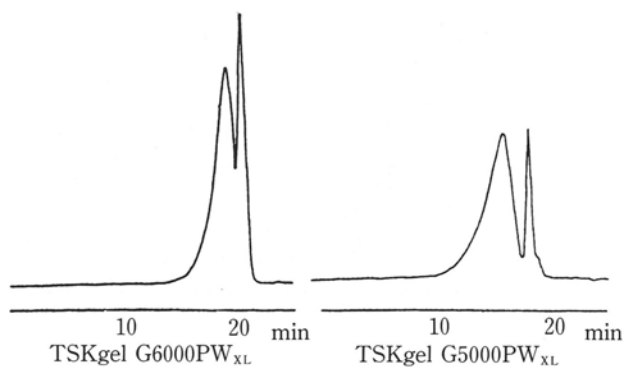
Water-soluble polymers				
Natural polymers		Semisynthetic polymers		Synthetic polymers
Starch	Sweet potato starch	Cellulose	Viscose	Polyvinyl alcohol
	Potato starch		Methylcellulose	
	Wheat starch		Ethylcellulose	Polyvinyl ether
			Hydroxyethyl cellulose	Polyvinylpyrrolidone
Mannan	Konnyaku		Carboxymethyl cellulose	Polyacrylamide
Seaweed	Glue plant (funori)	Starch	Soluble starch	Sodium polyacrylate
	Agar (galactan)		Carboxymethyl starch	Sodium polystyrene sulfonate
	Sodium alginate		Dialdehyde starch	Polyacrylamine
				Polyvinylpyridine hydrochloride
Plant mucilage	Abelmosk			Polyethylenimine
	Tragacanth gum,			
	Gum arabic			
Viscous substances derived from microbials	Dextran			
	Levan			
Proteins	Glue			
	Gelatin			
	Casein			
	Collagen			



**Figure 19 Separation of sodium polyacrylate**

Column size: 7.8 mm I.D. × 30 cm × 2

Eluent: 0.2 mol/L phosphate buffer (pH 6.9)



**Figure 20 Separation of DNA-Na**

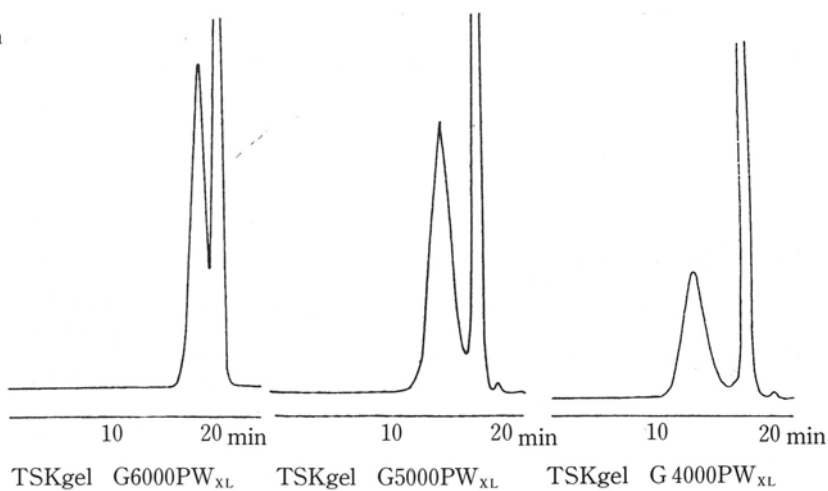
Column size: 7.8 mm I.D. × 30 cm × 2

Eluent: 0.2 mol/L phosphate buffer (pH 6.9)

**Figure 21 Separation of gelatin**

Column size: 7.8 mm I.D. × 30 cm × 2

Eluent: 0.2 mol/L phosphate buffer (pH 6.9)

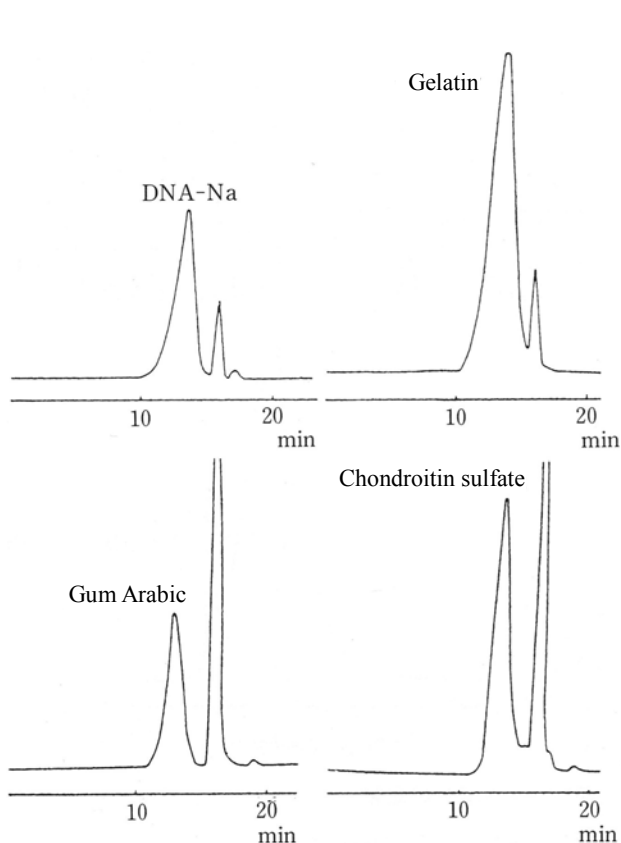


**Figure 22 Separation of chondroitin sulfate**

Column size: 7.8 mm I.D. × 30 cm × 2

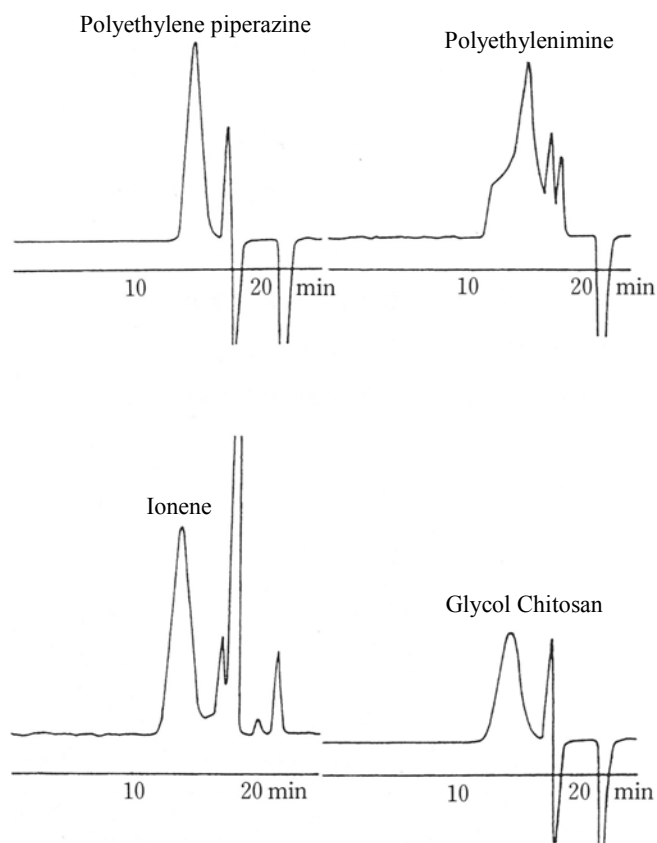
Eluent: 0.2 mol/L phosphate buffer (pH 6.9)

Figures 19-22 are chromatograms produced by analyzing the same samples using TSKgel G6000PW<sub>XL</sub>, TSKgel G5000PW<sub>XL</sub>, and TSKgel G4000PW<sub>XL</sub>, respectively. In Figure 23, the chromatogram shows the results of separation conducted using the TSKgel GMPW<sub>XL</sub>. Figure 24 shows the results of separation of polycations in an acetate buffer system using the TSKgel GMPW<sub>XL</sub>. Each of these chromatograms was produced at a flow rate of 1.0 mL/min with RI detection. See Separation Report No. 35 for further analytical details.



**Figure 23 Separation performed using TSKgel GMPW<sub>XL</sub> (1)**

Column size: 7.8 mm I.D. × 30 cm × 2  
 Eluent: 0.2 mol/L phosphate buffer (pH 6.9)



**Figure 24 Separation Performed using TSKgel GMPW<sub>XL</sub> (2)**

Column size: 7.8 mm I.D. × 30 cm × 2  
 Eluent: 0.5 mol/L phosphate buffer + 0.5 mol/L sodium acetate

**Table 8 Types of Eluents**

Type	Representative Eluent for use with PW
Non-electrolyte polymers and polyanions	0.2 mol/L phosphate buffer (pH 7) or 20 % CH <sub>3</sub> CN (or CH <sub>3</sub> OH) / 0.2 mol/L phosphate buffer (pH 7)
Polycations	0.5 mol/L acetate + 0.5 mol/L sodium acetate or 0.3 mol/L TEA + conc. phosphoric acid (pH 2.9)

TEA: Triethanolamine

## 10. Conclusion

The TSKgel PW<sub>XL</sub> series of columns provides much improved performance over conventional GPC columns. Use of these columns offers many advantages: separation time is reduced by half while achieving high peak height and high sensitivity; and because the quantity of solvent used is reduced by half, separation efficiency is improved.

Given that high-performance GPC satisfies 3 essential elements (ease of use, speed, and high sensitivity), this method of separation is expected to see increasing use in the future.